IMMUNIZATION BY INOCULATION OF DNA TRANSCRIPTION UNIT

Related Application

5   This Application is a Continuation-in-Part of U.S. Application Serial No. 08/009,833, filed January 27, 1993, now US PAT 5,643,578, which is a Continuation-in-Part of U.S. Application Serial No. 07/855,562 filed March 23, 1992, now abandoned. The teachings of these applications are incorporated herein by reference.

10   Government Support

15   Background of the Invention

Vaccination with inactivated or attenuated organisms or their products has been shown to be an effective method for increasing host resistance and ultimately has led to the eradication of certain common and serious infectious

20   diseases. The use of vaccines is based on the stimulation of specific immune responses within a host or the transfer of preformed antibodies. The prevention of certain diseases, such as poliomyelitis, by vaccines represents one of immunology's greatest triumphs.

25   Effective vaccines have been developed for relatively few of the infectious agents that cause disease in domestic animals and man. This reflects technical problems associated with the growth and attenuation of virulent strains of pathogens. Recently effort has been

30   placed on the development of subunit vaccines (vaccines that present only selected antigens from a pathogen to the

host). Subunit vaccines have the potential for achieving high levels of protection in the virtual absence of side effects. Subunit vaccines also offer the opportunity for the development of vaccines that are stable, easy to

5   administer, and sufficiently cost-effective for widespread distribution.

Summary of the Invention

This invention relates to a method of subunit vaccination. Specifically, this invention relates to a

10   method of immunizing an individual, comprising introducing into the individual a DNA transcription unit (or units) which comprises DNA encoding a desired antigen or antigens and DNA ~~encoging~~ encoding a transcriptional promoter element or elements. A single transcription unit or multiple DNA

15   transcription units can be administered to an individual to achieve immunization against one antigen or multiple antigens. The uptake of the DNA transcription units by host cells results in the expression of the desired antigen or antigens, thereby eliciting humoral or cell-

20   mediated immune responses or both humoral and cell-mediated responses. The elicited humoral and cell-mediated immune response can provide protection against infection by pathogenic agents, provide an anti-tumor response, or provide contraception. The host can be any

25   vertebrate, avian or mammalian, including humans.

The present invention relates to the use of DNA transcription units for raising immune responses. In one embodiment, the individual is immunized by parenteral routes of inoculation. These include intravenous,

30   intramuscular, intradermal, and subcutaneous administration of DNA transcription units. DNAs administered to the skin can be delivered with a DNA gun. In a second embodiment, the individual is immunized by contacting a mucosal surface, such as a respiratory

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mucosal surface, with DNA transcription units in such a manner that the transcription units are taken up by (i.e., enter the cells of) the mucosal surface. DNAs for mucosal administration can be microsphere encapsulated.

5   The DNA transcription units introduced by the present method can be used to express any antigen encoded by an infectious agent, such as a virus, a bacterium, a fungus, or a parasite, as well as antigenic fragments and peptides that have been experimentally determined to be effective

10   in immunizing an individual against infection by a pathogenic agent. As stated above, DNA transcription units can also be used for contraceptive purposes or for anti-cancer therapy.

The desired antigens to be expressed can be designed

15   so as to give internal, surface, secreted, or budding and assembled forms of the antigens being used as immunogens.

There are numerous advantages of the use of DNA for immunizations. For example, immunization can be accomplished for any antigen encoded by DNA. Furthermore,

20   the DNA encoded antigens are expressed as "pure" antigens in their native states and have undergone normal host cell modifications. Also, DNA is easily and inexpensively manipulated and is stable as a dry product or in solution over a wide range of temperatures. Thus, this technology

25   is valuable for the development of highly effective subunit vaccines.

Brief Description of the Drawings

Figure 1 is a schematic representation of a bacterial plasmid containing a DNA transcription unit (referred to

30   as

pP1/H7) comprising an influenza virus hemagglutinin type 7 (H7) gene expressed by a replication competent retroviral vector.

Figure 2 is a schematic representation of a bacterial plasmid containing a DNA transcription unit (p188) comprising an influenza virus hemagglutinin type 7 (H7) gene expressed by a replication defective retroviral

5 vector.

Figure 3 is a schematic representation of a bacterial plasmid comprising a retroviral vector (pRCAS) with no H7 insert, used as a control.

Figure 4A is a schematic representation of the

10 nonretroviral vector comprising the influenza virus antigen DNA transcription unit encoding subtype H7 hemagglutinin.

Figure 4B is a schematic representation of the nonretroviral vector comprising a control DNA

15 transcription unit, encoding no influenza virus antigens.

Figure 4C is a schematic representation of the nonretroviral vector comprising the influenza virus antigen DNA transcription unit encoding subtype H1 hemagglutinin.

20 Figure 5 is a bar graph depicting the cytotoxic T cell response of mice inoculated by gene gun with EDIM VP7 rotavirus cDNA in comparison with controls. Filled bars, effector to target ratio 60:1; striped bars, effector to target ratio 30:1.

25 Figure 6 is a schematic representation of the pJW4303 vector comprising the CMV intron A, a leader sequence for the tissue plasminogen activator (TPA) protein, and bovine growth hormone polyadenylation sequences.

Figure 7A is a schematic representation of HIV-1

30 proviral DNA.

Figure 7B is a schematic representation of the NL4-3.dpol insert.

Figure 7C is a schematic representation of the HXB-2.env insert.

Figure 7D is a schematic representation of the NL4-3.env insert.

Figure 8A is a schematic representation of HIV-1 env DNA.

5   Figure 8B is a schematic representation of a sgp120.env insert.

Figure 8C is a schematic representation of a sgp140.env insert.

Figure 8D is a schematic representation of a
10   sgp160.env insert.

Figure 9 is a representation of the DNA immunization schedule used in mice for HIV-1 vectors.

Figure 10 is a bar graph depicting the levels of anti-gp120 antibody raised in mice by intravenous and
15   intramuscular inoculations. Stippled bars, sera of those receiving NL4-3.env DNA; striped bars, NL4-3.dpol DNA; open bars, both NL4-3.env DNA and NL4-3.dpol DNA.

Figure 11 is a bar graph depicting the levels of anti-gp120 antibody raised in mice by intravenous and
20   intramuscular inoculations, followed by gene gun inoculations. Stippled bars, sera of those receiving NL4-3.env DNA; striped bars, NL4-3.dpol DNA; open bars, both NL4-3.env DNA and NL4-3.dpol DNA.

Figure 12 is a bar graph depicting the longevity of
25   mouse anti-gp120 titer in those animals receiving both NL4-3.env DNA and NL4-3.dpol DNA.

Figure 13 is a bar graph depicting the titration of anti-gp120 antibody raised in DNA inoculated mice. Stippled bars, sera of those receiving NL4-3.env DNA;
30   striped bars, NL4-3.dpol DNA; open bars, both NL4-3.env DNA and NL4-3.dpol DNA.

Figure 14 is a bar graph depicting levels of anti-env antibody raised in mice by gene gun env DNA inoculations. Light filled bars, results from a high responder mouse
35   inoculated only by gene gun; striped bars, results from a

moderate responder mouse inoculated only by gene gun; dark
filled bars, results from a mouse inoculated by
intravenous and intramuscular routes, with NL4-3.env,
bleed 8.

5   Figure 15 is a graph depicting the cytotoxic T-cell
activity of HIV-1 inoculated mice. Open circles, results
from mice inoculated with vector; closed circles, NL4-
3.env DNA; closed triangles, NL4-3.dpol DNA; closed
squares, both NL4-3.env DNA and NL4-3.dpol DNA. E:T =
10 effector to target ratio.

       Figure 16A is a schematic representation of SIV-239
proviral DNA.

       Figure 16B is a schematic representation of the
SIV239.dpol insert.

15   Figure 17A is a schematic representation of SIV-
239.env DNA.

       Figure 17B is a schematic representation of a SIV
sgp120 insert.

       Figure 17C is a schematic representation of a SIV
20 sgp140 insert.

       Figure 17D is a schematic representation of a SIV
sgp160 insert.

       Figure 18 is a schematic representation of the
restriction sites and oligonucleotides used for PCR
25 amplification and subcloning of envs from patient isolates
of HIV-1.

Detailed Description of the Invention

       This invention relates to a method of immunizing
vertebrates, particularly mammals, including humans,
30 against a protein, a pathogen, or an infectious agent,
thereby eliciting humoral and/or cell-mediated immune
responses which interfere with the activity of the
protein, or which limit the spread or growth of the
infectious agent and result in protection against

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subsequent challenge by the pathogen or infectious agent. In the method of the present invention, a DNA transcription unit is administered to an individual in whom immunization is desired.

5   The term "immunizing" refers herein to the production of an immune response in a vertebrate which protects (partially or totally) from the manifestations of infection (i.e., disease) caused by an infectious agent. That is, a vertebrate immunized by the present invention

10 will not be infected or will be infected to a lesser extent than would occur without immunization.

A DNA transcription unit is a polynucleotide sequence, bounded by an initiation site and termination site, that is transcribed to produce a primary transcript.

15 As used herein, a "DNA transcription unit" includes at least two components: antigen-encoding DNA and transcriptional promoter element or elements. Antigen-encoding DNA can encode one antigen or multiple antigens, such as multiple HIV antigens or antigens from two or more

20 different proteins or infectious agents. The DNA transcription unit can additionally be inserted into a vector which includes sequences for replication of the DNA transcription unit. A DNA transcription unit can optionally include additional sequences, such as: enhancer

25 elements, splicing signals, termination and polyadenylation signals, viral replicons and bacterial plasmid sequences. In the present method, a DNA transcription unit (i.e., one type of transcription unit) can be administered, or a combination of two or more types

30 of DNA transcription units can be administered.

The DNA transcription unit can be produced by a number of known methods. For example, using known methods, DNA encoding the desired antigen can be inserted into an expression vector. See Maniatis et al., Molecular

Cloning, A Laboratory Manual, 2d, Cold Spring Harbor
Laboratory Press (1989).

The DNA transcription unit can be administered to an
individual, or inoculated, in the presence of adjuvants or
5   other substances that have the capability of promoting DNA
uptake or recruiting immune system cells to the site of
the inoculation. It should be understood that the DNA
transcription unit itself is expressed in the host cell by
transcription factors provided by the host cell, or
10 provided by a DNA transcriptional unit.

The "desired antigen" can be any antigen or
combination of antigens expressed by an infectious agent,
or any antigen or combination of antigens that has been
determined to be capable of eliciting a protective
15 response. The desired antigen can also be a tumor antigen
or an antigen which provides protection against
conception. The antigen or antigens can be naturally
occurring, or can be mutated or specially modified. The
antigen or antigens can represent different forms, such as
20 subgroups (clades), subtypes or serotypes of an infectious
agent. These antigens may or may not be structural
components of a cell or an infectious agent. The encoded
antigens can be translation products or polypeptides. The
polypeptides can be of various lengths. They can undergo
25 normal host cell modifications such as glycosylation,
myristoylation or phosphorylation. In addition, they can
be designed to undergo intracellular, extracellular or
cell-surface expression. Furthermore, they can be
designed to undergo assembly and release from cells.

30   Potential pathogens for which the DNA transcription
unit can be used include DNA encoding antigens derived
from any virus, chlamydia, mycoplasma, bacteria, parasite
or fungi. Viruses include the herpesviruses,
orthomyxoviruses, rhinoviruses, picornaviruses,
35 adenoviruses, paramyxoviruses, coronaviruses,

rhabdoviruses, togaviruses, flaviviruses, bunyaviruses, rubella virus, reovirus, hepadna viruses and retroviruses including simian immunodeficiency virus or human immunodeficiency virus. Bacteria include mycobacteria,

5 spirochetes, rickettsias, chlamydia, and mycoplasma. Fungi include yeasts and molds. Parasites include malaria. It is to be understood that this list does not include all potential pathogens against which a protective immune response can be generated according to the methods

10 herein described.

An individual can be inoculated through any parenteral route. For example, an individual can be inoculated by intravenous, intraperitoneal, intradermal, subcutaneous or intramuscular methods, or by the gene gun.

15 An individual can be inoculated by any mucosal route. The DNA transcription unit can be administered to a mucosal surface by a variety of methods, including DNA-containing nose-drops, inhalants, suppositories or by microsphere encapsulated DNA. For example, the DNA transcription unit

20 can be administered to a respiratory mucosal surface, such as the nares or the trachea.

Any appropriate physiologically compatible medium, such as saline, is suitable for introducing the DNA transcription unit into an individual.

25 Immunization as described herein was accomplished with various DNA transcription units (e.g., vectors) that express different proteins. The DNA transcription units described herein are representative of the types of transcription units that can be used in the current

30 invention. The DNA transcription units can encode antigens from a single infectious agent, including antigens from different subgroups (clades) or subtypes of the infectious agent, and can additionally encode antigens from more than one infectious agent.

In one embodiment of the current invention, immunization was accomplished using a DNA transcription unit encoding the murine rotavirus neutralization capsid protein VP7. In another embodiment, influenza virus

5 hemagglutinin glycoproteins were used. The hemagglutinin glycoprotein mediates adsorption and penetration of virus and is a major target for neutralizing antibodies. Influenza virus hemagglutinin proteins have 14 different serological subtypes. In a particular embodiment, DNA

10 expression vectors for the H7 subtype (comprising a DNA transcription unit encoding the H7 subtype hemagglutinin) have been used to provide protection against challenge with an H7N7 virus in an avian model. In another particular embodiment, a DNA transcription unit expressing

15 the H1 hemagglutinin has been used to immunize against an H1N1 virus in a murine model. A mixture of DNA transcription units, comprising DNA encoding a variety of antigens from different subgroups (e.g., A and B) and/or from different subtypes (such as subtypes 1-14 of subgroup

20 A) of influenza, can also be used in the current invention.

In addition, experiments have been conducted to examine the immunogenicity of human immunodeficiency virus (HIV-1) and simian immunodeficiency virus (SIV_{mac}) DNA

25 transcription units in mice and monkeys. DNA vaccines for immunodeficiency virus are designed to raise broad cell-mediated as well as humoral responses. This is accomplished by using a mix of DNAs encoding different HIV antigens. This mix can include two main components. The

30 first, termed dpol constructs, serve to raise cytotoxic T-cell responses against a wide variety of HIV-1 proteins. The second, termed Env constructs, serve to raise antibody responses against the spectrum of Envs present in endemic infections.

The dpol DNA constructs encode a non-infectious DNA, because one of the nine proteins is not included in the construct. The constructs mimic many of the characteristics of a live infection. Attenuation can be

5   accomplished by a number of mutations. One example includes rendering the long terminal repeats (LTRs) non-functional by point mutations, deletions or truncations and rendering the polymerase gene non-functional by point mutations or internal deletions.

10   Altered expression of other genes is optional. Thus, these constructs give the opportunity for expressing 8 out of 9 HIV-1 proteins (Gag, Env, Vif, Vpr, Vpu, Tat, Rev and Nef) or 8 out of 9 SIV_{mac} proteins (Gag, Env, Vif, Vpr, Vpx, Tat, Rev and Nef). Expression of this extensive

15   repertoire of HIV-1 or SIV_{mac} proteins facilitates the activation of the cytotoxic arm of the immune system in individuals of diverse histocompatibility types. The number of epitopes expressed increases the chance that epitopes are present that can be recognized by individual

20   histocompatibility types. Each histocompatibility type will presumably recognize and present a subset of the expressed epitopes.

The dpol constructs also facilitate the raising of CTL responses that are directed against regulatory as well

25   as structural HIV-1 proteins. The expression of the regulatory proteins by the dpol DNAs affords the opportunity for raising cytotoxic responses against the proteins which are expressed earliest in infections and which are most active in latent infections. This offers

30   the vaccinated host the possibility of clearing cells before they have produced virus. It also affords the opportunity of clearing cells that have latent infections.

A mix of Env constructs is used in conjunction with the dpol constructs to raise a broad antibody response

against Env. A broad humoral response to Env is important because Env is the protein which is exposed on extracellular virus. Thus antibodies to Env can prevent infection by processes such as neutralization and
5 complement mediated lysis. A broad response to Env is important because Env is the HIV-1 protein which undergoes the most marked evolution in infected humans. The mix of Envs can include:

(1) Envs that represent different subgroups of HIV-1,
10   such as subgroups A to F. These are used to provide broad geographic protection (Myers et al., Human Retroviruses and AIDS, Los Alamos National Laboratory, Los Alamos NM (1992)).

(2) Envs from within a subgroup that represent growth
15   characteristics representative of different phases of infection or or have distinct tissue tropisms. These are used to provide protection against the spectrum of viruses present in an endemic infection.

(3) Envs representative of those favored for different
20   routes of transmission. These include Envs representative of homosexual transmission, heterosexual transmission, transmission by intravenous drug use, trans-placental transmission or neonatal transmission.

25 (4) Mutant forms of env that may be particularly effective at raising desired immune responses. An example of such an env is the HXB-2 Env that has been deleted for V1, V2 and V3 sequences by Dr. Joseph Sodroski (Dana Farber Cancer Institute, Boston, MA).
30   This Env retains the conformational epitopes associated with the CD4 binding domain and may be particularly effective in raising antibody that will have broad neutralizing activity (Wyatt, et al., 1993, J. Virol. 67:4557-4565).

Different structural forms of Env are expressed by
the DNAs used in vaccines. These forms can include a
soluble form of gp120, such as sgp120. This form is
particularly effective at raising antibody to V3 loop
5   determinants (Earl et al., 1994, J. Virol. in press). A
second form which can be expressed is a soluble form of
gp140, such as sgp140. This form represents an oligomer
of the extracellular components of Env; it raises antibody
against gp41 as well as antibody against gp120, and also
10 raises antibodies that recognize oligomeric forms of Env
that are present in the assembled envelope spikes found on
virions (Earl et al., J. Virol., in press; Moore, et al.,
J. Virol. 68:469-484 (1994)). The sgp120 and sgp140 are
released from cells, facilitating their entry into major
15 histocompatibility class II-dependent pathways of antigen
presentation and the raising of T-cell dependent B-cell
responses. A native form of Env (gp160) can also be
expressed. This membrane-anchored form of Env represents
the form of Env found on the surfaces of virions and
20 infected cells.

The following Examples describe vaccination trials
using direct DNA inoculations designed for use in
influenza virus, rotavirus, and immunodeficiency virus
models. Vaccination trials for influenza virus use avian,
25 murine, and ferret models. The avian and murine models
demonstrate protective immunizations against lethal
challenges, wherein challenge of an unimmunized animal
caused death. The ferret model demonstrated protective
immunizations against virus replication in the nares;
30 challenge of an unimmunized animal resulted in virus
replication in the nares. Vaccination trials for
rotavirus used a murine model. The murine model
demonstrated antibody and cytotoxic T-cell activity in
animals receiving DNA transcriptional units for rotavirus

protein, wherein animals receiving control DNA exhibited no antibody or cytotoxic T-cell activity for rotavirus. Vaccination trials for immunodeficiency virus used murine and simian models. The murine model demonstrated antibody including neutralizing antibody and cytotoxic T-cell activity in animals receiving DNA transcriptional units for human immunodeficiency virus type 1 (HIV-1), wherein animals receiving control DNA exhibited no antibody or cytotoxic T-cell activity. The simian model is used to test for the raising of responses that will protect against a lethal challenge with simian immunodeficiency virus-macaque (SIV_{mac}).

The current invention is illustrated by the following examples, which are not to be construed as limiting in any way.

Example 1 Immunization of Chickens Against Influenza Virus
A DNA transcription unit referred to as pP1/H7 (Fig. 1), encoding a replication competent avian leukosis virus expressing the influenza virus hemagglutinin type 7 (H7) gene was constructed as described in Hunt et al., J. of Virology, 62(8):3014-3019 (1988). DNA unit p188 (Fig. 2) encoding a replication defective derivative of pP1/H7 that expresses H7 but is defective for the avian virus vector polymerase and envelope proteins was constructed by deleting an XbaI fragment from pP1/H7. DNA unit pRCAS (Fig. 3), encoding the avian leukosis virus vector, with no influenza virus insert, was constructed as described in Hughes et al., J. of Virology, 61:3004 (1987). DNA units were diluted in saline at a concentration of 100 µg per 0.2 ml for inoculation.
To test the ability of the inoculated DNA to protect against a lethal influenza virus challenge, groups of

three-week old chicks were inoculated with pP1/H7, p188, or pRCAS DNA. Specific pathogen free chicks that are maintained as an avian-leukosis virus-free flock (SPAFAS, Norwich, CT) were used for inoculations. Each chick

5 received 100 μg of DNA intravenously (iv), 100 μg intraperitoneally (ip), and 100 μg subcutaneously (sc). Four weeks later, chicks were bled and boosted with 300 μg of DNA (100 μg iv, 100 μg ip, and 100 μg sc). At one week post-boost, chicks were bled and challenged by the nares

10 with 100 lethal doses$_{50}$ (1x10^4 egg infectious doses) of a highly pathogenic type H7 avian influenza virus, A/Chicken/Victoria/1/85 (H7N7) (Ck/Vic/85). The H7 gene of Ck/Vic/85 differs in about 15% of its codons from that of the immunizing H7 (Hunt et al., J. of Virology

15 62(6):3014-3019 (1988)). Thus, the Ck/Vic/85 challenge tests for the ability to achieve broad cross-protection within the H7 subtype. Ck/Vic/85 spreads rapidly throughout the internal organs and brain of chickens, causing death within 4-7 days. Post-challenge, the

20 chickens were observed daily for ten days for signs of disease. One and one half weeks after challenge, sera were obtained from surviving birds. These as well as the pre- and post-boost sera were used for analyses for anti-H7 antibodies (See below).

TABLE 1: Protection Against Lethal H7N7 Influenza Virus
with DNA Coding for H7 Hemagglutinin

		HI TITERS		
Group	# Survivors/ # Tested	Post-vaccine 4 weeks	Post-boost 1 week	Post-Challenge 1.5 weeks
pP1/H7	6/6	<.[a]	<.	864 (160-1280)
p188	6/6	<[b]	<	427 (160-1280)
pRCAS	0/6	<	<	+[c]

[a] (<.) means one of six birds had an HI titer of 10.
[b] (<) means that all birds had titers of less than 10.
[c] (+) means that all birds died.

10   The H7-expressing DNA transcription units protected
each of the chickens inoculated with pP1/H7 or p188 (Table
1). In contrast, inoculation with the control DNA, pRCAS,
failed to protect the chickens against lethal virus
challenge. The birds in the control group started to show
15 signs of disease on the second day post-challenge. By the
third day, three of the six control birds had died and all
control birds were dead by the fifth day. The birds
inoculated with hemagglutinin-expressing DNAs showed no
signs of disease. By one and one half weeks post
20 challenge both of these groups had developed high levels
of HI antibody.

        Additional Experiments
        To assess the reproducibility of the protection
elicited by immunization with the replication-defective
25 H7-expressing DNA, the experiment described above was
repeated three times using only p188 and pRCAS DNAs for
inoculations, with the difference that the chicks were
challenged two weeks post-boost instead of one week post-
boost as in the first experiment. Three-week-old SPAFAS

chicks were inoculated with 100 μg of DNA by each of three routes (iv, ip and sc). Four weeks later, they were boosted by inoculation with 100 μg of DNA administered iv, ip and sc. Two weeks later, chickens were challenged via
5   the nares with 100 lethal doses$_{50}$ of Ck/Vic/85 (H7N7).

The results of the repeat experiments confirmed that the H7-expressing p188 DNA could afford protection against a lethal challenge, as shown in Table 2.

TABLE 2:   Reproducibility of Protection Against a Lethal
10   H7 Virus Challenge by Immunization with p188
             DNA*

Fate of Challenge Group (# survivors/# tested)				
Expt.	p188 DNA	pRCAS DNA	Amantadine	No treatment
1	6/6	0/6	-	-
2	5/6	1/5	4/5	-
3	9/32	0/32	-	-
4	8/12	0/12	-	0/12
Total	28/56	1/55	4/5	0/12

* Experiment 1 is the same as that presented in Table 1.
20   -, not tested.

In contrast to the first experiment, in which all of the p188-inoculated chickens survived the lethal challenge, immunizations in the second, third, and fourth experiments resulted in protection in 28% to 83% of the vaccinated
25   birds. Further, in contrast to the first experiment in which vaccinated birds showed no signs of disease, most of the survivors of the repeat experiments showed transient signs of post-challenge sickness. As in the first experiment, the control DNA did not provide protection.
30   Summing the results of the four experiments, 28 out of 56 p188-vaccinated birds survived; only one of 55 control birds survived. Thus, highly significant protection was

achieved. This level of protection is particularly
impressive in view of the fact that immunizing and
challenge H7 genes had undergone antigenic drift of about
15% in the amino acid sequence.

5   Example 2 Analysis of Antibody Response to H7 in
                 Vaccinated and Unvaccinated Animals
         To allow the comparison of antibody responses to H7
     in vaccinated and unvaccinated chickens, Experiment 2 from
     Example 1 (see Table 2) included a non-vaccinated group
10   rescued with the anti-influenza A virus drug, amantadine-
     HCL (Webster, R.G., et al., J. Virol. 55:173-176 (1985)).
     All of the five amantadine-treated birds developed
     disease. Four of these survived, providing sera that
     could be used to compare antibody responses to H7 in
15   immunized and non-immunized chickens.
         Sera from p188 inoculated and amantadine-treated
     birds in the second experiment were analyzed for the time
     course of antibody responses to H7 and to other influenza
     virus proteins. Antibody responses to H7 were quantitated
20   using hemagglutination inhibition (HI) as well as virus
     neutralization and enzyme-linked immunosorbent assays
     (ELISA). In tests for hemagglutination inhibition, sera
     were analyzed in microtiter plates with receptor-
     destroying enzyme-treated sera as described by Palmer et
25   al., Advanced Laboratory Techniques for Influenza
     Diagnosis, p. 51-52, Immunology series no. 6, U.S.
     Department of Health, Education, and Welfare, Washington,
     D.C. (1975). Neutralizing antibody was determined in
     chick embryo fibroblast cultures. Neutralization tests
30   were for 200 $TCID_{50}$ of virus and used cytopathology and
     hemagglutination for detection of virus replication.
     Results are shown in Table 3, below.

TABLE 3: Antibody Response in p188-DNA-Immunized Compared with Amantadine-Rescued Birds

DNA	No.[a]	Bleed	Antibody to Ck/Vic/85 (H7N7)			Antibody to Ck/Penn/1370/83 (H5N2)
			HI	Neutralizing	ELISA (x10^{-3})	ELISA (x10^{-3})
p188	6	1 wk PB[b]	5 (0-10)	2 (0-10)	2 (0-10)	<
	6	2 wk PB	8 (0-20)	13 (0-33)	5 (0-10)	<
	5	1 wk PC[c]	112 (80-160)	873 (33-3333)	640 (100-1000)	26 (0-100)
	5	2 wk PC	272 (80-640)	540 (33-1000)	640 (100-1000)	46
None/ Aman.	5	1 wk PB	<[d]		<	<
	5	2 wk PB	<	<	<	<
	4	1 wk PC	<	<	<	<
	4	2 wk PC	300 (80-640)	442 (100-1000)	1000 (1000)	1000 (1000)

Antibody titers are given as the median (range).
[a] (No.) Number of chicks in group at time of bleed.
[b] (wk PB) means weeks post boost.
[c] (wk PC) means weeks post challenge.
[d] (<) means all birds had titers of less than 10.

Analysis of the antibody responses in vaccinated and amantadine-rescued birds revealed that the p188-inoculations had primed an antibody response to H7. As in experiment 1 (Table 1), DNA vaccination and boost induced

5 only low titers of antibody to H7. However, within one week of challenge, the DNA-immunized group had high titers of HI and neutralizing activity for H7. These titers underwent little (if any) increase over the next week. Furthermore, most of the post-challenge antibody in the

10 vaccinated birds was directed against H7. This specificity was shown by comparing ELISA antibody titers to H7 virus (the immunizing hemagglutinin type) and H5 virus (a hemagglutinin type to which the birds had not been exposed). The post-challenge sera contained 20-times

15 higher titers of ELISA antibody for the H7 than the H5 virus. By contrast, in the amantadine-rescued group, most of the antibody was not H7-specific and was directed against proteins that are common to the H5 and H7 virus. This was demonstrated by comparable titers of ELISA

20 antibody for the H5 and the H7 influenza viruses.

Example 3 Immunization of Chickens Using a Nonretroviral
          Transcription Unit

This experiment was performed in order to demonstrate that DNA transcription units devoid of retroviral DNA

25 could be successfully employed to generate a protective immune response according to the methods herein described. The vectors used in this experiment to vaccinate chickens are shown in Figures 4A and 4B. Figure 4A is a schematic representation of pCMV/H7, a plasmid capable of expressing

30 the influenza virus H7 subtype hemagglutinin under the transcription control of a cytomegalovirus (CMV) immediate early promoter. Figure 4B shows pCMV/control, a control plasmid which is not capable of expressing influenza antigens. These plasmids are derivatives of the pBC12/CMV

vector of Dr. Bryan Cullen, Duke University, Durham, North Carolina (Cullen, B.R., <u>Cell</u> <u>45</u>:973-982 (1986)).

In the experiments using pCMV/H7 (the nonretroviral-based DNA transcription unit) to generate immune

5　responses, immunization and boosts used the same inoculation schedule but different inoculation routes than described in Example 1. Specifically, 100 μg of DNA was inoculated by each of three routes: intravenous, intraperitoneal, and intramuscular. The boosts used the

10　same DNA dose and sites of inoculation as the vaccinations. Challenge was 1-2 weeks after the boost, with 100 lethal doses$_{50}$ of Ck/Vic/85 used so as to achieve essentially 100% killing. Results of five independent trials of pCMV/H7 DNA are shown in Table 4, below.

15　TABLE 4:　Protection Against a Lethal Ck/Vic/85 (H7N7)
　　　　　　　Influenza Virus Challenge by Immunization with
　　　　　　　pCMV/H7 DNA

| Trial | Fate of challenge Group (# survivors/# tested) | |
	pCMV/H7 DNA	pCMV/Control DNA
1	5/6	0/6
2	4/6	0/6
3	2/6	0/7
4	4/6	1/7
5	4/6	0/7
Total	19/30	1/33

25　　　　In the five chicken trials using pCMV/H7 for vaccination, approximately 60% of the chickens were protected. This level of survival was very similar to that achieved with the retroviral-based vector p188 (Table 2). As in the trials with the retrovirus-based vector,

30　many of the survivors developed transient signs of

influenza. Thus, protection comparable to that achieved with the retrovirus-based vector could be achieved with a non-retroviral based vector.

Antibody responses to H7 in experiments with the non-
5 retroviral-based vector were also similar to those with the retroviral-based vector (Table 3, Table 4). Specifically, protective responses were associated with the rapid rise (within one week) of H7-specific antibodies after challenge. Sera contained low to undetectable
10 levels of anti-H7 antibodies after vaccination and boost. The low levels of antibody prechallenge coupled with the rapid rise in antibody post challenge suggest that protection was mediated by the DNA transcriptional units having established memory responses that were mobilized by
15 the challenge infection. Thus, considerable protection has been achieved using nonretroviral DNA expression vectors (containing DNA transcription units encoding viral antigens) to vaccinate animals.

Example 4 Immunization of Mice by Vaccination with pCMV/H1
20 **DNA: Analysis of Various Routes of Inoculation**
A DNA transcription unit referred to as pCMV/H1 was successfully used to immunize mice against a lethal challenge with mouse adapted A/PR/8/34 H1N1 influenza virus. This transcription unit encodes an influenza type
25 H1 hemagglutinin under the transcriptional regulation of the CMV immediate early promoter. The H1 influenza virus hemagglutinin gene used in this construct is described in more detail in Winters et al., Nature 292:72 (1981) and is identical to that in the challenge virus. The vector is
30 the same as that used in Example 3 for the expression of H7 (See Fig. 4B). The pCMV/H1 transcription unit is depicted in Figure 4C.

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TABLE 5: Antibody Responses to the H7 Challenge Virus in pCMV/H7 and pCMV-control DNA inoculated chickens

Time of Bleed	Trial	Control DNA-inoculated			CMV/H7 DNA/inoculated		
		HI	Neutral-izing	ELISA (x10^{-3})	HI	Neutral-izing	ELISA (x10^{-3})
4 weeks post-vaccination (pre-boost)	2	<	<	<	<	<	<
	3	<	<	<	<	<	<
	4	<	<	<	<	<	<
	5	<	<	<	2.5	<	<
1 week post-boost (pre-challenge)	2	<	<	<	<	<	<
	3	<	<	<	<	<	<
	4	<	<	<	2.5	<	2.5
	5	<	<	<	2.5	<	2.5
2 week post-challenge	2	dead	dead	dead	60	33	765
	3	dead	dead	dead	60	33	1000
	4	dead*	dead*	dead*	100	33	775
	5	dead	dead	dead	140	108	1000

* One control bird survived in this group. Its post challenge titers were HI, 80; Neutralizing, 10, and ELISA, 100.

Vaccine trials in mice were accomplished by the administration of DNA to 6- to 8-week-old BALB/c mice. Two DNA inoculations were given, one at time 0 and the second four weeks later. Test animals were observed

5 throughout the trials and mice were weighed regularly beginning at the time of challenge. Lethal challenge was administered at 10 days after the second inoculation by inhalation of virus into the lungs of Metofane (Pitman-Moore, Mundelein, IL)-anesthetized mice. The challenge

10 consisted of 250 plaque-forming units (10-100 times the median lethal dose (LD_{50})) of mouse-adapted A/PR/8/34 (H1N1) influenza virus in 100 μl of saline supplemented with 0.1% bovine serum albumin. The challenge virus underwent localized replication in the respiratory tract

15 causing death due to pneumonia within 1-2 weeks. Routes of DNA inoculation included the following: intravenous (tail vein); intraperitoneal; intramuscular (both quadriceps); intranasal (DNA drops administered to the nares of mice anesthetized with Metofane); intradermal

20 (foot pad); and subcutaneous (scruff of the neck). In general, 100 μg of DNA was administered in 100 μl of saline per test site. For foot-pad inoculations, 50 μg of DNA was administered in 25 μl.

Table 6 sets forth the results showing protection of

25 the mice against a lethal A/PR/8/34 (H1N1) influenza virus challenge by inoculation of pCMV/H1 DNA in saline. Data in Table 6 are pooled from four independent trials. Routes shown are intravenous (iv), intraperitoneal (ip), intramuscular (im), intranasal (in), intradermal (id), and

30 subcutaneous (sc). Signs of influenza included weight loss, ruffled fur, and lethargy. The signs were scored as follows: +, transient weight loss but maintenance of smooth fur and normal levels of activity; ++, transient weight loss, some ruffling of fur and lethargy; +++,

transient weight loss and more severe ruffling of fur and
lethargy; ++++, more pronounced weight loss coupled with
severe ruffling of fur and lethargy; +++++, weight loss
and severe signs of influenza leading to death.

5 Probability was calculated by using Fisher's exact two-
tailed test comparing the frequency of survival and
mortality in vaccine versus control groups.

Table 6:   Protection against Lethal A/PR/8/34 (H1N1)
                  Challenge by Various Routes of pCMV/H1
10   Inoculation

DNA	Route	Dose (μg)	Signs of Infl.	# Surv. /# Test	% Surv	Proba-bility
pCMV/H1	iv,ip,im	300	++	21/22	95	<0.0001
	im	200	++	18/19	95	<0.0001
	iv ~~in~~	100	++	10/12	83	<0.0001
	in	100	+++	13/17	76	<0.0001
	id	50	++++	9/12	75	<0.001
	sc	100	++++	4/6	67	<0.02
	ip	100	+++++	0/6	0	
pCMV/ control	various	0-300	+++++	3/24	13	

15   Excellent survival occurred in groups receiving
intramuscular inoculations, intravenous inoculations, or
inoculations by each of three routes (intramuscular,
intravenous, and intraperitoneal). The relatively mild
influenza that developed in these groups was associated
20 with ruffling of fur and transient weight loss. Good
survival, but more severe influenza, occurred in mice
receiving DNA intranasally. Yet poorer survival (67-75%)
and more severe signs of influenza occurred in mice
receiving intradermal and subcutaneous inoculations. None

of the mice receiving only intraperitoneal injections survived the lethal challenge. Control groups (inoculated with pCMV/control DNA or no DNA) developed severe signs of influenza with very few mice (13%) surviving the

5 challenge. Thus, intramuscular, intravenous, intranasal and intradermal routes of administration each provided good protection.

These results indicate that the efficiency of transfection does not necessarily determine the efficiency

10 of vaccination. The high ability of rodent muscle to take up and express DNA (Wolff, J.A. et al., _Science_ _247_:1465-1468 (1990); Wolff, J.A. et al., _BioTechniques_ _11_: 474-485 (1991); and Acsadi, G. et al., _New Biol._ _3_:71-81 (1991)) did not correlate with an unusual efficiency of

15 intramuscular vaccinations. Intramuscular inoculations worked well, but no better than, intravenous inoculations, and only somewhat better than the administration of DNA nose drops. Similar results were obtained in chicken trials for route of inoculation (described below), where

20 intravenous and intratracheal inoculations achieved levels of protection comparable with those provided by intramuscular inoculations.

Example 5 _Protective Immunizations by Different Routes of_ _pCMV/H7 Inoculation in the Chicken Influenza_

25 _Model_

The effect of the route of inoculation on DNA vaccination was further evaluated in the avian influenza virus model. The DNA vaccine trials for route of inoculation in chickens were conducted as described in

30 Example 1. Routes of inoculation included the following: intravenous (wing vein): intramuscular (breast muscle): intratracheal (DNA drops administered to the trachea): subcutaneous (nape); intrabursal (injections just above the chicken's vent); and intraorbital (DNA drops

administered to the eye). In general, 100 or 200 μg of
DNA was administered in 200 μl of saline. Results are
shown in Table 7, below. Data for p188 and pRCAS DNA are
pooled from three independent trials. Data for pCMV/H7
5 and pCMV/control DNA are pooled from two independent
trials. Probability was calculated by using Fisher's
exact two-tailed test comparing the frequency of survival
and mortality in vaccine versus control groups. Routes
shown are intravenous (iv), intraperitoneal (ip),
10 subcutaneous (sc), intramuscular (im), intratracheal (it),
intrabursal (ib), and intraorbital (io).

Table 7:   Protection of Chickens against a Lethal Ck/
           Vic/85 (H7N7) Challenge Following Different
           Routes of DNA Inoculation[a]

DNA	Route	Dose (μg)	# Surv. /# test	% Surv.	Proba-bility
	iv	100	8/33	24	<0.01
	ip	100	0/8	0	
	sc	100	0/2	0	
pRCAS	iv,ip,sc	300	1/55	2	
	im	200	3/10	30	<0.02
	it	200	3/10	30	<0.02
	ib	200	1/10	10	
	io	200	1/10	10	
pCMV/ control	Various	0-300	1/43	2	

[a] Data for controls is the same as in Tables 2 and 4.

20   In the chicken trials for route of inoculation, good
efficacy was demonstrated for intravenous, intramuscular,
and mucosal administration of the vaccine DNAs.   These
groups exhibited about one half the level of protection
(24-30%) achieved in groups receiving DNA by each of three

routes (Tables 2 and 4). Poor to no protection was achieved by subcutaneous, intraperitoneal, intrabursal, and intraorbital inoculations. Chickens receiving control DNA developed lethal influenza, with very few chickens

5 (approximately 2%) surviving the challenge. Within experimental groups, surviving chickens showed variability in the severity of influenza-related illness.

Example 6 Protective Immunizations by Gene Gun Delivery of DNA to the Mouse Epidermis

10   To test whether a DNA gun could be used to deliver DNA to the epidermis, the Accell particle bombardment device (Agracetus, Middleton, WI) was employed to deliver DNA-coated gold beads to the epidermis of mice. These experiments were done in collaboration with Dr. Joel R.

15 Haynes of Agracetus, Inc.

        For gene-gun delivery of DNA to mice, plasmid DNA was affixed to gold particles by adding 10 mg of 0.95 μm gold powder (Degussa, South Plainfield, NJ) and an appropriate amount of plasmid DNA to a 1.5-ml centrifuge tube

20 containing 50 μl of 0.1 M spermidine. Plasmid DNA and gold were coprecipitated by the addition of 50 μl of 2.5 M $CaCl_2$ during vortex mixing, after which the precipitate was allowed to settle and was washed with absolute ethanol and resuspended in 2.0 ml of ethanol. The gold/DNA

25 suspension was transferred to a capped vial and immersed in a sonicating water bath for 2-5 seconds to resolve clumps. The 163 μl of the gold/DNA suspension was layered onto 1.8 cm x 1.8 cm Mylar sheets and allowed to settle for several minutes, after which the meniscus was broken

30 and excess ethanol was removed by aspiration. Gold/DNA-coated mylar sheets were dried and stored under vacuum. The total amount of DNA per sheet was a function of the DNA/gold ratio and ranged from 0.2 to 0.0002 μg per sheet.

Animals were anesthetized with 30 μl of Ketaset/Rompun
(10:2). Abdominal target areas were shaved and treated
with Nair (Carter-Wallace, New York) for two minutes to
remove residual stubble and stratum corneum. Target areas
5   were thoroughly rinsed with water prior to gene delivery.
DNA-coated gold particles were delivered into abdominal
skin with the Accell instrument, which employs an electric
spark discharge as the motive force (Yang, M.S. et al.,
Proc. Natl. Acad. Sci. USA 87: 9568-9572 (1990)). Each
10 animal received two nonoverlapping deliveries per
immunization, at a discharge voltage of 17 kV. The beads
deliver DNA into cells, where the DNA dissolves and can be
expressed (Yang, M.S. et al., Proc. Natl. Acad. Sci. USA
87: 9568-9572 (1990); Williams, R.S. et al., Proc. Natl.
15 Acad. Sci. USA 88: 2726-2730 (1991)). Expression is
transient, with most of the expression being lost within
2-3 days due to the normal sloughing of the epidermis
(Williams, R.S. et al., Proc. Natl. Acad. Sci. USA 88:
2726-2730 (1991); unpublished observations).
20   Gene gun-based acceleration of DNA-coated gold beads
into the epidermis proved to be by far the most efficient
method of DNA immunization, as shown in Table 8. Data are
pooled from four independent trials. Probability was
calculated by using Fisher's exact two-tailed test
25 comparing the frequency of survival and mortality in
vaccine versus control groups. For the description of the
signs of influenza, see above discussion of Table 6.

Table 8: Protection against Lethal A/PR/8/34 (H1N1)
Challenge by Gene Gun-Delivered pCMV/H1 DNA
Inoculation

DNA	Dose (μg)	Signs of Infl.	# Surv./ # Test.	% Surv.	Proba-bility
pCMV/H1	0.4	+	21/22	95	<0.0001
	0.04	+++	7/11	64	<0.001
	0.004	+++++	0/5	0	
	0.0004	+++++	0/4	0	
pCMV/ control	0.4	+++++	3/22	14	

These tests of gun-delivered DNA in the murine model
demonstrated that as little as 0.4 μg of DNA was
sufficient to achieve 95% survival. These survivors
developed very limited to no signs of postchallenge
influenza. Mice receiving 0.04 μg of gun-delivered
pCMV/H1 DNA had an approximately 65% survival rate and
suffered fairly severe signs of influenza. Mice that
received 0.004 μg or 0.0004 μg of pCMV/H1 DNA succumbed to
the challenged. As in tests of saline injections, mice
receiving control DNA developed severe signs of influenza
and had very limited survival (14%). Thus, highly
efficient immunizations were achieved by gene-gun delivery
of DNA to the epidermis of mice. This method of
immunization required 250-2500 times less DNA than the
saline inoculations (0.4-0.004 μg as opposed to 100-200 μg
of DNA) (See Tables 6 and 7).

Example 7 Antibody Responses in pCMV/H1 Vaccinated Mice

In the murine trials described above in Examples 4
and 6, sera were collected immediately prior to each DNA
inoculation, immediately prior to challenge, and at two
times after challenge. Anesthetized mice were bled from

the eye vein into 40 μl nonheparinized microhematocrit
tubes. Sera for members within a group were pooled.
Hemagglutination inhibition assays were performed with
chicken red blood cells and mouse serum that had been
5   pretreated with kaolin to remove background activity
(Novak, M. et al., Vaccine 11: 55-60 (1993)).
Hemagglutination inhibition titers are the reciprocal of
the highest serum dilution giving complete inhibition of
hemagglutination. The isotypes of mouse antibodies were
10   determined by enzyme-linked immunosorbent assay (ELISA)
using standard protocols and microwell plates coated with
purified A/PR/8/34 (H1N1) influenza virus. These assays
used 1:1000 dilutions of isotype-specific peroxidase-
conjugated antibodies that had been provided at titers
15   with similar activities (Sigma Immuno-Chemicals). Data
for serum analysis are shown below in Table 9. Data are
the geometric means of the reciprocal of the final
dilutions of pooled sera that scored positive for a given
condition.

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Table 9: Antibody Responses in Vaccine Trials Testing
Routes of pCMV/H1 DNA Inoculation in Mice

			Titers of Antibody to A/PR/8/34 (H1N1)			
				ELISA value x10^{-2}		
Route	Bleed	# Test.	HI	IgM	IgG	IgA
pCMV/H1 in saline						
iv	Prevac	2 (12)	<	<	<	<
	10 d PB	2 (12)	<	<	8	4
	4 d PC	1 (6)	20	<	8	4
	14-19 d PC	2 (10)	113	1	256	4
im	Prevac	3 (19)	<	<	<	<
	10 d PB	3 (19)	<	<	3	<
	4 d PC	2 (13)	6	<	32	2
	14-19 d PC	3 (18)	127	<	406	2
in	Prevac	3 (17)	<	<	<	<
	10 d PB	3 (17)	<	<	2	1
	4 d PC	2 (11)	<	1	2	1
	14-19 d PC	3 (17)	160	2	202	2
pCMV/control in saline						
various	Prevac	3 (16)	<	<	<	<
	10 d PB	3 (16)	<	<	<	<
	4 d PC	2 (9)	<	<	<	<
	14-19 d PC	1 (2)	320	<	256	<
pCMV/H1						
gene gun	Prevac	2 (10)	<	<	<	<
	10 d PB	3 (16)	10*	1	10	<
	4 d PC	3 (16)	20*	2	64	<
	14-19 d PC	3 (15)	160*	2	64	<
pCMV/control						
gene gun	Prevac	2 (12)	<	<	<	<
	10 d PB	3 (16)	<	1	<	<
	4 d PC	3(16)	<	2	<	<
	14-19 d PC	1 (3)	NT	4	512	<

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* Only one of the three pools of sera was tested for HI
activity. Prevac = bleed before DNA vaccinations; 10 d
PB; bleed 10 days after the second DNA inoculation (prior
to challenge); 4 d PC, bleed 4 days post challenge. #
5 Test. = the number of groups for which pooled sera were
assayed (total number of animals contributing sera to the
pools). NT = not tested. < = activity not detected in the
lowest dilution of serum used in tests.

       DNA vaccinations by the various routes appeared to
10 prime memory antibody responses. The DNA vaccinations and
booster inoculations raised only low to undetectable
titers of hemagglutination-inhibiting antibodies and ELISA
activity. These low levels of activity underwent rapid
increases after challenge. As in the chicken experiment
15 (Tables 3 and 5) protection occurred in animals that did
not have detectable levels of anti-influenza antibodies
before challenge. However, the best protection occurred
in groups in which the DNA inoculations had raised
detectable titers of prechallenge antibody.
20   Use of ELISAs to score the isotypes of the anti-
influenza virus antibodies demonstrated that the
immunizations had primed IgG responses. Low titers of
anti-influenza IgG could be detected prechallenge in the
sera of mice vaccinated by gun delivery, intravenous or
25 intramuscular inoculations of DNA. Borderline to
undetectable titers of IgG were present in the sera of
mice receiving DNA nose drops (consistent with the poorer
protection provided by this route of DNA administration).
By four days after challenge, increased levels of IgG were
30 detected in mice undergoing the best protection. By
contrast, mice receiving control DNA did not have
detectable levels of anti-influenza virus IgG until the
second serum collection after challenge. This was
consistent with vaccinated, but not control, groups
35 undergoing a secondary antibody response to the challenge.

These experiments show that the DNA inoculation had primed both T-helper and B-cell memory. This memory appeared to provide protection by supporting the mounting of secondary responses in challenged animals. Evidence for

5 the priming of memory is provided by the DNA inoculations raising antibodies belonging to the IgG isotype, as IgG is produced by differentiated plasma cells that have undergone immunoglobulin rearrangements in response to T-cell help (Abbas, A.K. et al., Cellular and Molecular

10 Immunology (Saunders, Philadelphia, PA), pp. 187-197 (1991)). Evidence for the mobilization of memory in response to the challenge is found in the rapid increases in serum IgG after challenge.

Marginal to undetectable levels of IgM and IgA were

15 detected in both prechallenge and postchallenge sera. The low levels of these immunoglobulin isotypes throughout the trials indicated that none of the routes of DNA inoculation were effective at raising serum IgM or IgA.

Example 8 Use of pCMV/H1 DNA Transcriptional Unit to

20   Protect Ferrets Against A/PR/8/34(H1N1)
             Influenza Challenge

Studies on pCMV/H1 DNA immunization in a ferret model were undertaken because this influenza model has many similarities to human influenza infections. In the

25 initial experiment, ferrets were immunized with purified pCMV/H1 DNA in saline by intramuscular inoculations at a one month interval. Young adult female ferrets were prebled and vaccinated with 500 μg of pCMV/H1 or pCMV/control DNA in saline by two injections of 125 μl in

30 each hind leg for a total inoculation volume of 500 μl. One ferret received three intramuscular inoculations of 500 μg of pCMV/H1 DNA at one month intervals while a second animal received two intramuscular inoculations of 500 μg of DNA at one month intervals. The control animal

received three 500 μg intramuscular inoculations of
pCMV/control DNA at one month intervals.

Metofane-anesthetized ferrets were challenged with
$10^{7.7}$ egg infectious doses$_{50}$ of A/PR/8/34 (H1N1) via the
5   nares at one week after the final DNA inoculation. Nasal
washes were collected at days 3, 5 and 7 post challenge
under ketamine anesthetic. Titration of virus in nasal
washes was done in eggs as described (Katz, J.M. and R.G.
Webster, *J. Infect. Dis. 160*: 191-198 (1989)). Data are
10   presented in Table 10, below.

**TABLE 10: Protection of Ferrets against an H1 Virus by
Intramuscular Inoculation of pCMV/H1 DNA**

DNA	No. of DNA Adminis-trations	Ferret ID No.	Virus Titer in Nasal Washes, $\log_{10}$ egg infectious doses$_{50}$/ml		
			day 3	day 5	day 7
pCMV/H1	3	901	5.5	1.5	<1
	2	903	5.7	4.7	<1
pCMV/control	3	907	6.5	6.2	<1

Analyses of nasal washes revealed similar high titers
of virus in the washes of all of the ferrets at 3 days
post challenge. Interestingly, the ferret receiving three
inoculations of pCMV/H1 had largely cleared the nasal
20   infection by five days post challenge, with its five day
nasal wash containing less than 10 egg infectious doses$_{50}$
of virus per ml. At this time the ferret receiving two
inoculations of pCMV/H1 DNA had a ten fold reduction in
the titer of virus in its nasal wash. By contrast, the
25   ferret receiving control DNA had modest if any reduction
in the titer of virus in its nasal wash. By 7 days post
challenge, all of the ferrets had cleared their nasal
infections. The much more rapid clearing of virus in the

ferret receiving three intramuscular inoculations of
pCMV/H1 DNA and the somewhat more rapid clearing of virus
in the ferret receiving two intramuscular inoculations of
pCMV/H1 DNA than in the two ferrets receiving control DNA
5   suggest that the intramuscular inoculations of pCMV/H1 had
raised some anti-influenza immunity.

Gene Gun Inoculation
     To increase the efficiency of the induction of
immunity, a second experiment was undertaken in ferrets
10   using the Accell gene gun to deliver DNA coated gold beads
into the skin of ferrets. The abdominal epidermis was
used as the target for gene gun delivered DNA with ferrets
receiving two gene gun administrations of DNA at a one
month interval. Gene gun inoculations were delivered to
15   Ketamine-anesthetized young adult female ferrets. Skin
was prepared by shaving and treating with the depilatory
agent NAIR (Carter-Wallace, New York). DNA beads (1 to 3
microns) were prepared for inoculations as previously
described (Fynan et al., Proc. Natl. Acad. Sci. USA
20   90:11478-11482 (1993)). A delivery voltage of 15 kV was
used for inoculations. Ferrets were inoculated with
either 2 μg or 0.4 μg of DNA. Ferrets inoculated with 2
μg of DNA received 10 shots with each shot consisting of
0.8 mg of beads coated with 0.2 μg of DNA. Ferrets
25   receiving 0.4 μg of DNA received two of these shots.
     Metofane-anesthetized ferrets were challenged at one
week after the second DNA immunization by administration
of $10^{6.7}$ egg infectious doses of A/PR/8/34 (H1N1) virus via
the nares. This challenge was 10 fold lower than in the
30   experiment using intramuscular inoculation because of the
high levels of virus replication in the first challenge.
Nasal washes were collected at days 3 and 5 post challenge

under ketamine anesthetic and the virus titered as
described above. Data are presented in Table 11, below.

Table 11: Protection of Ferrets against an H1 Virus by
Gene Gun Inoculation of pCMV/H1 DNA

DNA	Amount of DNA (μg)	Ferret ID No.	Virus Titer in Nasal Washes, $\log_{10}$ egg infectious doses$_{50}$/ml	
			day 3	day 5
pCMV/H1	2	927	<1	<1
		931	<1	<1
		933	<1	<1
	0.4	926	4.3	<1
		929	3.9	<1
		933	<1	<1
pCMV/control	2	932	3.5	<1
		934	3.7	<1

Analyses of post-challenge nasal washes in gene gun
vaccinated ferrets revealed that the three ferrets
receiving 2 μg of DNA and one of the three ferrets
receiving 0.4 μg of DNA were completely protected from the
challenge. This was shown by the inability to recover
virus in the nasal washes of these animals at 3 days post
challenge. The remaining two animals receiving 0.4 μg of
DNA and the control animals were not protected, with
easily detected titers of virus present in the nasal
washes of the animal at three days post challenge. In
this experiment, all animals (control and vaccinated) had
no detectable virus in their nasal washes by five days
post challenge.

Ferrets from the gene gun experiment were next
analyzed for antibody responses to the DNA administrations
and to the challenge virus. These assays tested for

neutralizing activity for A/PR/8/34 (H1N1). The
titrations of antibodies were done as described (Katz,
J.M. and R.G. Webster, *J. Infect. Dis. 160*: 191-198
(1989)). Titers of neutralizing activity are the
5   reciprocals of the highest dilution of sera giving
complete neutralization of 200 50% tissue culture
infectious doses of virus. Data are presented in Table
12, below.

Table 12: Neutralizing Antibody in Ferrets Vaccinated with Gene Gun-Delivered pCMV/H1 DNA and Challenged with A/Pr/8/34 (H1N1) Influenza Virus

DNA	Amount of DNA (μg)	Ferret ID No.	Neutralizing Antibody			
			Pre-inoculation	Post-boost, pre-challenge	Post challenge (7 days)	Post challenge (14 days)
pCMV/H1	2	927	<10	<10	251	178
		931	<10	79	251	178
		933	<10	13	398	398
	0.4	926	<10	<10	2511	2511
		929	<10	<10	398	126
		933	<10	<10	794	562
pCMV/control	2	932	<10	<10	562	398
		934	<10	<10	562	794

Neutralizing antibody post DNA boost but prior to challenge was detected in two of the animals receiving 2 μg of gene gun-delivered DNA. No neutralizing antibody was detected in the pre-challenge sera of the third animal

5 receiving 2 μg of DNA (an animal that was completely protected against the presence of virus in nasal washes). Neutralizing antibody was also not detected in the sera of the ferret receiving 0.4 μg of DNA that did not develop virus in its nasal wash.

10 In animals with prechallenge antibody, protection was presumably due to the presence of neutralizing antibody as well as the mobilization of memory responses for neutralizing antibody. In protected animals without detectable levels of prechallenge antibody, protection was

15 likely due to the rapid mobilization of memory responses by the infection, with the mobilized responses controlling the infection. Protection in vaccinated animals in the absence of prechallenge antibody has also been observed in prior DNA vaccination studies in mice and chickens (see

20 Tables 3, 5 and 9) (Fynan et al., Proc. Natl. Acad. Sci. USA 90:11478-11482 (1993); Robinson et al., Vaccine 11: 957-960 (1993)) and in vaccine trials using retrovirus and pox virus vectors to express the influenza virus hemagglutinin glycoprotein (Hunt et al., J. Virol.

25 62:3014-3019 (1988); Webster et al., Vaccine 9: 303-308 (1991)).

Example 9 Microsphere-encapsulated DNA for Mucosal
        Administrations

The mucosal route of DNA inoculation was further

30 developed by testing for the ability of microsphere-encapsulated DNA to raise protective responses against a lethal influenza virus challenge. The murine influenza virus model was used for these studies. pCMV/H1 and pCMV/control DNA were encapsulated in alginate

microspheres at the Virus Research Institute, Inc.,
Cambridge, MA. A trial was conducted in which each group
received a primary inoculation and a boost. Group A
received 0.4 μg of gene gun delivered DNA for the primary

5   and no boost. Group B received 0.4 μg of gene gun
delivered DNA for the primary and the boost. Group C
received 0.4 μg of gene gun delivered DNA for the primary
and 100 μg of alginate-encapsulated DNA for the boost.
Group D received 100 μg of alginate-encapsulated DNA for

10 both the primary and boost inoculations. Each
administration of alginate-encapsulated DNA was delivered
in 100 μl of water to the nares of Metofane-anesthetized
mice. Lethal challenge with 500 pfu of A/PR/8/34 (H1N1)
influenza virus was administered via the nares to

15 metofane-anesthetized mice at 10 days after the second DNA
inoculation. Four control groups received pCMV/control
DNA at the same amounts and following the same regimen as
the groups receiving pCMV/H1 DNA. Data for this
experiment are presented in Table 13.

Table 13: Protection of Mice Against a Lethal Influenza
Virus Challenge by Administration of Vaccine DNA
in Microspheres

		pCMV/H1 DNA		pCMV/Control DNA	
DNA	Boost	Signs of Infl.	Surv./ Total	Signs of Infl.	Surv./ Total
0.4 μg, gun*	none	+++	3/6	++++	2/6
0.4 μg, gun	0.4 μg, gun	-	6/6	+++++	1/6
0.4 μg, gun	100 μg, ms i.n.	++	5/6	++++	2/6
100 μg, ms i.n.	100 μg, ms i.n.	+	4/6	++++	1/4

*gun = gene gun delivery; ms i.n. = DNA encapsulated in
alginate microspheres

The intranasal administration of alginate-encapsu-
lated DNA provided good protection. Each of the vaccine
groups receiving alginate-encapsulated pCMV/H1 DNA
exhibited much better survival than the groups receiving
alginate-encapsulated pCMV/control DNA. Four out of 6 of
the mice receiving only alginate-encapsulated DNA survived
with very modest signs of influenza. By contrast, only
one out of 4 mice receiving alginate-encapsulated control
DNA survived. All of the control group developed severe
signs of influenza. The group receiving only one gun
inoculation exhibited moderate signs of influenza and had
a 50% survival rate (3/6 mice). Addition of an alginate
boost provided better protection against signs of
influenza and a higher survival rate (5/6 mice surviving).
Two gene gun deliveries of DNA provided the best survival,
with all of the mice surviving with no signs of influenza.

Example 10   <u>Immunization of Mice Using a DNA</u>
<u>Transcription Unit Encoding a Rotavirus</u>
<u>Protein</u>

A rotavirus DNA transcription unit was tested for its
5   ability to immunize mice. The pCMV/VP7 vector used in
this experiment to vaccinate mice for rotavirus is similar
to those shown in Figure 4A and 4C, except that the
plasmid pCMV/VP7 is one capable of expressing the murine
rotavirus neutralization capsid protein VP7. VP7 DNA was
10 obtained from Dr. Harry Greenberg, Stanford University,
Palo Alto, CA, USA.

In the mouse experiments using pCMV/VP7 to generate
immune responses, 0.4 μg of DNA was gene gun delivered to
abdominal skin (as described above). All vaccinations
15 were followed by a boost 4 weeks later. The boosts used
the same DNA dose and sites of inoculation as the
vaccinations. Testing for antibody and cytotoxic T cells
(CTL) was 1-2 weeks after the boost. Data are shown in
Table 14 and Figure 5.

Table 14: Anti-VP7 Antibody in Sera of pCMV/VP7 DNA Inoculated Mice

Inoculation	OD_{492nm} (X ± SD, n = 3)*			
	Preinoculation Sera		Immune Sera	
	1:50	1:200	1:50	1:200
pCMV/VP7-gun	0.041 ± 0.040	0.000 ± 0.000	0.430 ± 0.220	0.130 ± 0.185
pCMV-gun	0.050 ± 0.070	0.050 ± 0.058	0.000 ± 0.000	0.044 ± 0.052
EDIM-p.o.	0.000 ± 0.000	0.000 ± 0.000	0.629 ± 0.220	0.589 ± 0.184

*Anti-VP7 antibody was tested by ELISA against whole mouse EDIM rotavirus.

The antibody ELISA titers against whole EDIM virus raised by pCMV/VP7 DNA are shown in Table 14. Antibody titers of 1:200 were raised by the DNA transcriptional unit for the VP7 gene (pCMV/VP7). The titer of antibody

5   obtained by one inoculation of live EDIM murine rotavirus gave a titer of 1:800 (not shown). No significant titer was obtained by the pCMV/control plasmid alone.

It was also found that the plasmid pCMV/VP7 was able to induce a cytotoxic T cell (CTL) response against murine

10 rotavirus infected cells. The results of a chromium-release CTL assay are shown in Figure 5. The percent specific lysis in spleen cells from mice inoculated with pCMV/VP7 was approximately 30% at an effector to target ratio of 60:1, compared to 45% lysis obtained with mice

15 orally infected with EDIM rotavirus.

Example 11   DNA Constructs for Immunization Against
                HIV-1

Two series of DNA transcription units are prepared for immunizations against HIV-1. The first of these uses

20 the pBC12/CMV vector (See above and Fig. 4B) to provide transcriptional control elements for HIV-1 sequences. In pBC12/CMV vectors, HIV-1 protein expression is Rev dependent. The second series uses the JW4303 vectors developed at James I. Mullins laboratory (Stanford

25 University) (Palo Alto, CA) (see Fig. 6). These vectors support Rev-independent expression of Env. The JW4303 vectors and accompanying oligonucleotides are designed to facilitate the cloning of PCR amplified fragments of the Env of any isolate of HIV-1.

30   pBC12/CMV Based Vectors

All cloning into the pBC12/CMV based vectors was performed in pBC12/CMV/IL2. Specifically, insert fragments were substituted for the BamHI to HinDIII

fragment of IL-2 cDNA. Three inserts have been used in these clonings (Figs 7B-7D).

pCMV/HIV-1-NL4-3.dpol (NL4-3.dpol) (Fig. 7B)

5   Figure 7A depicts HIV-1-NL4-3 (NL4-3) proviral DNA and its associated long terminal repeats (LTR) sequences and open reading frames. pNL4-3 was provided by Dr. Malcolm A. Martin's Laboratory (National Institutes of Health, Bethesda, MD) (Adachi, et al., J. Virol. 59:284-291 (1986)). The Genbank accession number for
10 strain HIV.NL4-3 is M1991. NL4-3.dpol inserts were constructed to encode non-infectious HIV-1 particles and to mimic a live but non-infectious infection. To achieve an insert that would encode non-infectious particles, the entire 5' and most of the 3' LTR were deleted from pNL4-3
15 using HaeII and BanII digestions respectively. The pol gene was rendered non-functional by a 1932 bp internal BalI deletion. Western blot analyses of transfected Cos cells were used to demonstrate the expression of Gag and Env. Gag and Env proteins were present both in cells and
20 in culture medium. This was anticipated because Gag is the only HIV-1 protein required for particle formation.

pCMV/HIV-1-HXB-2.env (HXB-2.env) (Fig. 7C)

HXB-2.env was designed to express complete HXB-2 Env and Rev. The Genbank accession numbers for strain
25 HIV.HXB2 are K03455 and M38432. Rev was included in the construct because expression of the normal HIV-1 Env is Rev dependent. This construct was achieved by substituting the SalI to XhoI fragment of the pSVIII.env construct of Dr. Joseph Sodroski (Dana Farber Cancer
30 Institute, Boston, MA) (Helseth, et al., J. Virol. 64:2416-2420 (1990)) for the BamHI to HindIII fragment of IL-2 in pBC12/CMV/IL-2. Western blot analyses of transfected Cos cells demonstrated the expression of Env.

pCMV/HIV-NL4-3.env (NLV-3.env) (Fig. 7D)

A second example of a construct expressing HIV-1 Env and REV, NL4-3.env, expressed HXB-2/NL4-3 Env fusion proteins and Rev. In this construct, unique restriction
5 sites near the ends of the HXB-2 env, KpnI and BamHI, were used to substitute NL4-3 sequences for homologous HXB-2 env sequences in pCMV/HXB-2.env. Western blot analyses of transfected Cos cells demonstrated the expression of Env.

JW4303 Based Vectors

10 The JW4303 plasmid uses ~2000 bp from the CMV immediate early promoter and sequences from the bovine growth hormone for insert expression (Fig. 6). Sequences from the CMV immediate early promoter include sequences encoding the CMV intron A. This intron can enhance the
15 expression of inserted genes (Chapman, et al., Nucleic Acids Research 14:3979-3986 (1991)). The JW4303 vector includes a synthetic leader sequence for the tissue plasminogen activator (TPA) protein. This synthetic leader provides the start site for Env expression. The
20 tissue plasminogen activator leader facilitates synthesis and secretion of glycosylated proteins (Haigwood, et al., Prot. Eng. 2:611-620 (1989)). PCR amplification from designer oligonucleotides is used to create env fragments that are inserted in-frame with the TPA leader. Consensus
25 5' oligonucleotides for sequences in different subgroups of HIV-1 allow the fusion of any HIV-1 env sequences to the TPA leader at or near the normal end of the mature Env. The oligonucleotides are designed to allow the use of unique restriction sites in the synthetic TPA leader
30 for subcloning into JW4303. For example, the 5' oligonucleotide JApcr503 includes an XbaI site that allows fusion of the TPA leader just prior to amino acid 6 of the mature Env for subgroup B isolates of HIV-1.

4 8

Three types of antisense oligonucleotides allow
construction of secreted gp120 (sgp120), secreted gp140
(sgp140), or a normal gp160 form of Env (see Figs. 8A-8D).
DNA fragments encoding sgp120 forms of Env are synthesized
5 using consensus antisense oligonucleotides at or near
sequences encoding the proteolytic cleavage site between
gp120 and gp41. An example of such an oligonucleotide for
subgroup B virus is JApcr504 (SEQ ID NO. 2). DNA encoding
sgp140 forms of Env are synthesized using an antisense
10 oligonucleotides for consensus sequences at or near the
membrane anchor domain of gp41. An example of such an
oligonucleotide for subgroup B viruses is JApcr502 (SEQ ID
NO. 3). DNAs encoding complete Envs are synthesized using
oligonucleotide encoding consensus sequences within the
15 cytoplasmic domain or 3' to the C-terminus of gp160. An
example of such an oligonucleotide for subgroup B HIV-1 is
JApcr506. The antisense oligonucleotides have unique
restriction sites that facilitate cloning into JW4303 or
derivatives of JW4303. For example JApcr502 and JApcr504
20 contain BamHI sites for cloning into the unique BamHI site
in JW4303.

JW4303/HIV-1-HXB-2.sgp120 (HXB-2.sgp120)

HXB-2.sgp120 (See Fig. 8B) was synthesized using
JApcr503 (gtcgctcctctagattgtgggtcacagtctattatggggtacc)
25 (SEQ ID NO. 1) and JApcr504
(ggtcggatccttactgcaccactcttctctttgcc) (SEQ ID NO. 2) to
amplify sequences from pCMV/HXB-2.env. The amplified
fragments were digested with XbaI and BamHI and subcloned
into NheI and BamHI digested JW4303. Western blot
30 analyses of transfected Cos cells revealed sg120 in both
the transfected cells and the culture medium.

JW4303/HIV-1-HXB-2.sgp 140 (HXB-2.sgp140)

HXB-2.sgp140 (See Fig. 8C) was constructed using JApcr503 (SEQ ID NO. 1) and JApcr502 (cgacggatccttatgttatgtcaaaccaattccac) (SEQ ID NO. 3) to
5   amplify sequences from pCMV/HXB-2.env. The amplified fragments were digested with XbaI and BamHI and subcloned into NheI and BamHI digested JW4303. Western blot analyses of transfected Cos cells revealed sg120 in both the transfected cells and the culture medium.

10 **Example 12   Immunogenicity Tests of HIV-1-NL4-3**
                    pBC12CMV Based Vectors

A trial immunization with the pBC12/CMV based vectors was conducted in BALB/c mice. Six to 8 week old mice were immunized with a series of injections of 200 μg of DNA
15   administered by both the intravenous (iv) and intramuscular (im) routes (the immunization schedule is shown in Fig. 9). Four experimental groups of six mice received DNA. Group A received pCMV/control DNA without an insert. Group B received pCMV/NL4-3.env DNA. Group C
20   received pCMV/NL4-3.dpol DNA. Group D received a mixture of 100 μg of pCMV/NL4-3.env and 100 μg of pCMV/NL4-3.dpol DNA. Mice were bled prior to immunization and at various times after immunization. At each bleed, sera from the mice present in a test group were pooled. At the end of
25   the experiment, mice were sacrificed and the spleens harvested for assays for cytotoxic T-cell (CTL) activity against a defined CTL epitope in the NL4-3 Env for BALB/c mice.

Levels of anti-Env antibody were scored using enzyme
30   linked immunoadsorbent assays (ELISA). Wells of a microtiter plate were coated with 0.4 μg of purified gp120 per well (purchased from American Biotechnology Inc., Cambridge, MA). Mouse sera were pretreated with kaolin to remove non-specific activity (Novak, et al., Vaccine

11:55-60 (1993)). Different dilutions of test sera were incubated in the wells and the amount of anti-gp120 IgG scored using alkaline phosphatase or horse radish peroxidase conjugated goat anti-mouse IgG. Appropriate
5   substrates were added and color development evaluated using an ELISA reader to determine optical densities. Optical density values obtained for sera of the control group (group A) were subtracted from values obtained for experimental groups (groups B,C,D).
10   The DNA immunizations raised long-lived antibody responses to the HIV-1 envelope (Figs. 10, 11 and 12). Easily detected levels of anti-Env antibody appeared by bleed 4 (1.5 weeks after the third DNA immunization) (Fig. 10). These levels of antibody persisted, with similar
15   levels of antibody present at bleed 5 (4 weeks post the third DNA incubation) (Fig. 10). The clustered DNA immunizations 4, 5 and 6 substantially raised the titer of anti-Env antibody (see bleed 8 at 1.5 weeks after immunizations 4, 5 and 6) (Fig. 10). These higher levels
20   of antibody persisted, exhibiting only a slow decline with time. Two gene gun inoculations to the abdominal epidermis of 0.4 μg DNA per each of two shots did not increase the titer of anti-Env antibody (Fig. 11). Experimental animals were maintained for an additional 14
25   weeks after the gene gun inoculations. During this time good titers of anti-Env antibody persisted (Fig. 12).
        pCMV/NL4-3,env DNA, pCMV/NL4-3.dpol DNA and the mixture pCMV/NL4-3.env and pCMV/NL4-3.dpol DNA had overall similar abilities to raise antibody (Fig. 10). For each
30   of these DNAs, sera with the highest levels of anti-Env antibody were obtained at bleed 8 (Figs. 10 and 11). Each of these sera had end point liters of about 1:3200 (Fig. 13). The mixture of dpol and env DNA raised the highest liters of anti-Env IgG (Figs. 10, 11 and 13). However,
35   these higher titers were less than 4-fold different from

those raised by the DNAs given as single species (Fig. 13).

The levels of anti-Env ELISA antibody raised by the multiple DNA inoculations in saline and a gene gun boost
5 were compared to those obtained by Joel Haynes at Agracetus, Inc. (Middleton, WI) using only gene gun delivery of an Env-expressing DNA (Fig. 14). The levels present in the pooled sera from bleed 8 for NL4-3.env inoculated mice from the experiment in Figure 9 were
10 intermediate between those found in the sera of a high responder mouse and a moderate responder mouse receiving only gene gun DNA. This was consistent with the sera that had been pooled from test groups in the experiment showin in Figure 9 representing sera with titers much like those
15 raised by three gene-gun deliveries of 0.4 µg DNA to the abdominal epidermis.

Sera from mice were also examined for neutralizing activity to NL4-3. These tests were conducted by incubating 50 to 100 infectious units of NL4-3 with
20 various dilutions of mouse sera that had been heat inactivated and Kaolin treated. Incubations were conducted for one hour at 37°C. Following the incubation, exponentially growing H9 cells were added. 24 hours later, the cells were washed and fed with fresh medium.
25 At four days the cultures were lysed with Triton-X100 and analyzed for the replication of NL4-3 using an antigen capture ELISA. The results of three such assays are summarized in Table 15. The data in Table 15 are the reciprocals of the last dilution of sera giving ≥90%
30 inhibition of NL4-3 replication. HIV-Ig is pooled immunoglobulin from sero-positive humans obtained from the AIDS repository, Bethesda, MD.

Table 15: Titer of Neutralizing Antibody for NL4-3 in DNA-
Raised Sera

Animal #	Control	Bleeds 7,8,9	Bleed 11	Bleed 13	HIV-Ig
313	1:40	>1:1000			
322	1:80	1:5120	1:5120	1:5120	
324	<1:100	1:6400	1:6400	1:1600	1:100

The neutralization tests revealed excellent
neutralizing activity in the DNA-immunized mice. In
agreement with the ELISA data, mice immunized with N4L4-
3.env and NL4-3.dpol DNA had comparable titers of
neutralizing antibody. Also in agreement with the ELISA
data, the neutralizing antibody exhibited excellent
persistence, undergoing ≤ a four fold drop in the 14 weeks
following the last DNA inoculation. Thus the DNA
inoculations had raised outstanding neutralizing activity
against NL4-3. This reflects the presentation of native
forms of HIV-1 Env by DNA-expressing cells.

Tests for cytotoxic T-cell (CTL) activity were
carried out by Dr. Joel Haynes, Agracetus, Inc. For CTL
analyses, mice were sacrificed and responder splenocytes
harvested and resuspended in RPMI 1640, 10% fetal calf
serum, 50 μg/ml gentamicin (RPMI-10) containing 10 units
of rat interleukin-2 per ml. Stimulator splenocytes were
prepared by suspending splenocytes from naive animals in
RPMI-10 at a concentration of 1x10^7 cells per ml and
adding mitomycin C to a final concentration of 25 μg per
ml. Stimulator cells were incubated in the presence of
mitomycin C for 25 minutes at 37°C, washed with RPMI-10,
and then pulsed with a synthetic peptide representing a
known CTL epitope recognized by BALB/c mice
(RIQRGPGRAFVTIGK) (SEQ ID NO. 4). Roughly equal numbers
of stimulator and responder cells were cocultured for 5 to

6 days. A cytotoxicity assay was employed to measure the ability of the in vitro stimulated responder cells to lyse chromium^{51} loaded peptide pulsed BALB/c 3T3 target cells.

The DNA immunizations raised cytotoxic T-cell
5 activity that was readily detected at the termination of the experiment (14 weeks after the last DNA immunization) (Fig. 15). CTL activities for Env peptide-pulsed target cells were similar for mice immunized with pCMV/NLA-3.env DNA, pCMV/NL4-3.dpol DNA and the mix of pCMV/NL4-3.env and
10 pCMV/NL4-3.dpol DNA.

Example 13   DNA Constructs for Immunization against
**   SIV$_{mac}$**

SIV Constructs

As with HIV-1, two series of DNA transcription units
15 have been prepared for immunizations against SIV$_{mac}$. The first of these uses the pBC12/CMV vector of Dr. Bryan R. Cullen (see above and Fig. 4B). The second series used the JW4303 vectors developed at James I. Mullins laboratory (See above and Fig. 6).

20   **pBC12/CMV Based SIV Vectors**

Cloning into the pBC12/CMV based vectors was performed in pBC12/CMV/IL-2. SIV239 inserts were prepared from plasmids encoding SIV239 proviral DNA (Fig. 17A). These plasmids (p239SpSp5' and p239SpE3') were provided by
25 Dr. Ronald C. Desrosiers, New England Regional Primate Research Center (Southborough, MA). Specifically a pCMV/SIV239.dpol (239.dpol) insert (Fig. 17B) was substituted for the BamHI to HindIII fragment of IL-2 cDNA in pBC12/CMV/IL-2. The 239.dpol insert was constructed by
30 rendering the 5' LTR non-functional by a NarI deletion, rendering pol nonfunctional by an internal BstEII deletion, and removing most of the LTR with a StuI

digestion. p239SpE3′ encodes a defective nef gene, indicated by the stipple. Western blot analyses of transfected Cos cells were used to demonstrate the expression of Gag and Env. Gag and Env proteins were

5   present both in cells and in culture medium. This was anticipated because Gag is the only SIV-1 protein required for particle formation.

JW4303 Based Vectors

The JW4303 DNA transcription units for SIV were

10 constructed using PCR amplified fragments of SIV env sequences (Figs. 17A-17D). A 5′ sense primer supported construction of a DNA encoding a fusion protein with the TPA leader. 3′ antisense oligonucleotides were used to create a sgp120, a sgp140 and full length SIV env

15 fragments.

JW4303/SIV239.sgp120 (239.sgp120) (Fig. 17B)

239.sgp120 was synthesized using oligonucleotide JApcr19 and oligonucleotide JWpcr8 to amplify sequences from p239spE3′. The amplified fragments were digested

20 with NheI and BamHI and subcloned into NheI and BamHI digested pJW4303. Western blot analyses of transfected Cos cells revealed sg120 in both the transfected cells and the culture medium. 239 sequences were used in this construct because SIV 239 represents an established model

25 for vaccine trials in macaques. SIV 239 is a mutant form of SIV 251, which is also used to generate constructs (see below). The Genbank accession numbers for strain SIV239 are M33262, M61062, and M61093. The Genbank accession numbers for strain SIV251 are M19499 and X06393.

pJW4303/SIV239.sgp140 (239.sgp140) (Fig. 17C)

30

239.sgp140 was constructed using oligonucleotides JApcr19 and HKpcr2 to amplify sequences from p239SpE3′.

The amplified fragments were digested with NheI and BamHI and subcloned into NheI and BamHI digests pJW4303. Western blot analyses of transfected Cos cells revealed sg140 in both the transfected cells and the culture

5 medium. As indicated above, 239 sequences were used because the 239 virus is established as a model for vaccine trials in macaques.

pJW4303/SIV251.sgp140 (251.sgp140) (Fig. 17C)

251.sgp140 was constructed using oligonucleotides

10 Japcr19 and Hkpcr2 to amplify sequences from pM40KSIV251env (obtained from Dr. Ronald C. Desrosiers, New England Primate Research Center, Southborough, MA). The amplified fragments were digested with NheI and BamHI and subcloned into NheI and BamHI digests JW4303. Western

15 blot analyses of transfected Cos cells revealed sgp140 in both the transfected cells and the culture medium.

pJW4303/SIV316.sgp140 (316.sgp140) (Fig. 17C)

316.sgp140 was constructed using oligonucleotides Japcr19 and Hkpcr2 to amplify sequences from PCR env clone

20 316-3 obtained from Dr. Ronald C. Desrosiers, New England Primate Research Center. The amplified fragments were digested with NheI and BamHI and subcloned into NheI and BamHI digested pJW4303. Western blot analyses of transfected Cos cells revealed sg120 in both the

25 transfected cells and the culture medium. SIV 316 is a mutant form of SIV 239 which has a monocyte/macrophage tropism, (Mori, K. et al., J. Virology 66(4):2067-2075 (1992)).

Example 14   SIV DNA Vaccine Trial Design

30   A vaccine trial was undertaken using SIV-encoding DNAs to immunize Rhesus macaques. Young male and female immunocompetent animals are used in the trials. Three

clusters of DNA inoculations are given at 1 and 3, 11 and 13, and 21 and 23 weeks of the trial. A lethal challenge is administered two weeks after the final DNA inoculation. This challenge consists of 10 monkey infectious units of

5   SIV239 administered by intravenous inoculation.

Three groups of monkeys have been placed in the trial. Each group is receiving three different SIV239 DNAs: 239.dpol, 239.sgp120, and 239.sgp140 (Figs. 16D, 17B and 17c). At each DNA inoculation, the first group of

10   four macaques is receiving 500 μg of each of these DNAs by both iv and im routes of inoculation as well as 2 gene gun shots (Accell Instrument) of each of the three DNAs to the thigh skin and two gun shots of each of the three DNAs to the abdominal skin. The second group of monkeys is

15   receiving two gene gun shots of each of the three DNAs to thigh skin and two gun shots of each of the DNAs to abdominal skin. The third group is receiving 500 μg of the pCMV/control DNA and 1 mg of the pJW4303 DNA by both intravenous and intramuscular routes of inoculation as

20   well as two gene gun shots of the pCMV/control and four gun shots of the pJW4303 DNA administered to the thigh skin and two gene gun shots of the pCMV/control and four gun shots of the pJW4303 DNA administered to abdominal skin.

25   Gene gun shots of 239.dpol have been accomplished with beads loaded with equimolar amounts of DNA transcriptional unit for SIV$_{mac}$ Rev and 239.dpol. The expression of additional Rev in skin cells increases the level of Gag and Env expression. The transcriptional unit

30   for SIV$_{mac}$ Rev was obtained from Dr. Gregory A. Viglianti, University of Massachusetts Medical Center, Worcester, MA.

Additional Env-encoding DNAs are added to the vaccine for inoculation at weeks 11 and 13 and 21 and 23. These are added to broaden the immune response to include

responses against SIV mutants that arise in infected animals. To accomplish the broadening of the response, the two vaccine groups of monkeys (groups one and two above) receive two gene gun shots of 251.sgp140 and two gene gun shots of 316.sgp140. These are delivered to the abdominal epidermis. These shots are given in addition to the same shots received at weeks one and three of the trial.

Example 15   Molecular Cloning of HIV-1 env Sequences
                  from Patient Isolates for Use in Vaccine
                  Transcription Units

To obtain env sequences for subgroup B HIV-1 isolates representing the slow/low, non-syncytium inducing, monocyte/macrophage tropic viruses characteristic of the healthy seropositive phase of infection as well as env sequences representing the rapid/high, syncytium inducing T-cell line tropic viruses found in patients with AIDS, two series of serial patient isolates were obtained from Dr. Eva Maria Fenyo, Karolinska Institute, Sweden (Von Gegerfelt, A. et al., Virol. 185: 162-168 (1991)) (see Table 16). These isolates were obtained over a two to three year period of time during the progression from the healthy, sero-positive phase, to the AIDS phase of infection. One series was from patient 5, and the second from patient 6.

Table 16: Designations and Biological Characteristics of Patient Isolates to be Used as a Source of Vaccine DNAs

Isolate	CD4+ cells/μl	Replication in PBLs	Cell lines	Syncytium Formation	Susceptibility to Neutralization	
					homologous	heterologous
5A	487	slow	no	-		
5B	226	rapid	yes	+++	B+,C+	1+,2+
5C	100	rapid	yes	+++	B-,C-	1+,2-
6A	370	slow	no	-		
6B	470	slow	no	-/+		
6C	450	slow	tran*	++	A+,B-,C+,D+	1+,2+
6D	197	rapid	yes	+++	A-,B-,C-,D-	1-,2+

* tran = transitory. Sera and isolates harvested at the same time are indicated by the same letters. For example, isolate 5B was obtained at the same time as serum B from patient 5. Heterologous sera represent sera from two patients with good neutralizing activity.

Envelope sequences from the isolates have been recovered by polymerase chain reaction (PCR) amplification. Culture supernatants were grown once on mitogen stimulated PBLs. DNA was prepared at five days post

5 infection when high levels of syncytia had appeared for the more virulent isolates. A nested PCR amplification was then used to recover a KpnI to BamIII fragment of env with molecular weight of approximately 2.1 kb. This fragment encodes essentially all of gp120, lacking codons

10 for only the 13 N-terminal amino acids of gp120. It also encodes approximately 240 of the approximately 340 amino acids of gp41, including all of the extracellular and transmembrane domains of gp41 as well as the portion of the intracellular domain that includes a highly acidic

15 sequence.

Oligonucleotide primers for PCR reactions were chosen to include conserved restriction endonuclease sites as well as to maximize the number of 3' bases that are completely conserved among the current HIV-1 isolates in

20 Myers data base (Myers et al., Human Retrovirus and AIDS, Los Alamos National Laboratory, Los Alamos, NM, 1992). Actual amplifications have used 250 ng of DNA, 25 pmole of each primer, and one unit of Amplitaq in a final volume of 100 μl.

25 Clones are cloned into a right hand half of pNL4-3 provided by Dr. Ronald C. Desrosiers, New England Primate Research Center, Southborough, MA). This is performed as a threepiece cloning of (i) a 2.1 kb KpnI to BamHI fragment of PCR amplified env sequences; (ii) a 0.6 kb

30 EcoRI to KpnI fragment from pNL4-3 (3' vpr to 5' env sequences); and (iii) the EcoRI to BamHI framgnet of the right half of pNL4-3 (contining 3' env, nef, LTR and plasmid sequences). As clones are obtained, a preliminary V3 loop sequence is obtained (see Table 17). This

35 sequence verifies that the clones are not a contaminant

and provides a signature sequence for monitoring further
subcloning.

Table 17: env Clones from Serial Isolates from Patients 5 and 6, and Preliminary V3 Loop Sequence

Isolate	Clone	pNL4-3/ env	V3 Loop	SEQ ID NO.
5A	EMF		CTRPNYTTRKRIHIGPGRAFYTTKNIIGNIKQAH	5
5B	4B-2	p5B-1	CTRPNYKTRSRIHIGPGRAFYTTKNIRGDIRQAHC	6
	EMF		CTRPNYKTRSRIHIGPGR	7
5C	EMF		CTRPNYKTSRRIHIGPGRSFYT	8
6A	4A-27	p6A-1	CTRPNNNTRKSIHIGPGRAIYTTGQIIGDIRQAHC	9
	EMF		CTRPNNNTRKSIHIGPGRAIYTT	10
6B	EMF		CTRPNNNTRKSIHIGPGRAFYTT?AIIGDIR	11
6C	A4-2		CTRPNNNTRRRIHIGPGRAIYTTGQIIGDIRQAHC	12
	A4-4		CTRPNNNTRRRIHIGPGRAIYTTGQIIGDIRQAHC	13
	A4-5		CTRPNNNTRRRIHIGPGRAIYTTGQIIGDIRQAHC	14
	4C-1	p6C-1		15
	EMF		CTRPNNNTRRRIHIGPGRA	16 /15
6D	4A-64		CTRPNNNTRRRIHIGPGRAIYTTGQIIGDIRQAHC	17 /16
	4D-6	p6D-1	CTRPNNNTRRRIHIGPGRAIYTTGQIIGDIRQAHC	18 /17
	EMF		CTRPNNNTRRRIHIGPGRA	19 /18
Consensus B			CTRPNNNTRKSIHIGPGRAFYTTGEIIGDIRQAHC	20 /19

EMF = Fenyo lab.

Clones are then tested for biological activity by co-transfection with the left half of pNL4-3 into COS-1 cells and co-cultivation with mitogen stimulated PBLs. The p6B-1, p6A-1, p6C-1 and p6D-1 NL4-3.env recombinants encode
5 functional envs that support the growth characteristics of the stocks from which they were recovered. These envs represent different stages of disease and different growth characteristics of patient isolates. The envs are moved on PCR amplified fragments into the pJW4030 vector for
10 immunogenicity tests (see above, Figs. 8A-8D).

Equivalents

Those skilled in the art will recognize, or be able to ascertain, using no more than routine experimentation, many equivalents to the specific embodiments of the

5 invention described herein. These and all other such equivalents are intended to be encompassed by the following claims.